

Five New C₁₉-Diterpenoid Alkaloids from *Aconitum hemsleyanum*

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Five new C₁₉-diterpenoid alkaloids, named hemsleyaconitines A – E (**1** – **5**, resp.), were isolated from *Aconitum hemsleyanum* PRITZ. By UV, IR, MS, 1D- and 2D-NMR analyses, their structures were elucidated as 18-dehydroxygeniculatine D (**1**), 6-hydroxy-14-O-veratroylneoline (**2**), 14-O-acetyl-8-ethoxysachaconitine (**3**), 18-veratroylkarakoline (**4**) and 8-O-ethylaustroconitine B (**5**).

Introduction. – *Aconitum hemsleyanum* PRITZ. (Ranunculaceae), a perennial herb distributed in Sichuan and Yunnan Provinces of China, has long been used as a folk medicine to treat rheumatism and pains [1]. Previous studies on this plant revealed that aconitine-type C₁₉-norditerpenoid alkaloids were the main constituents [2 – 4]. During our search for bioactive chemicals from natural sources, *A. hemsleyanum* was phytochemically investigated to afford five new C₁₉-norditerpenoid alkaloids, as well as four known ones, sachaconitine [5], karakoline [6], acoforestinine [7], and isotalatizidine [8], identified by comparing the spectral data with those reported. All of the compounds **1** – **5** (Fig. 1), as well as sachaconitine, karakoline, and acoforestinine, showed positive reactions to Dragendorff's reagent. Here, we report the isolation and structure elucidation of the five new alkaloids.

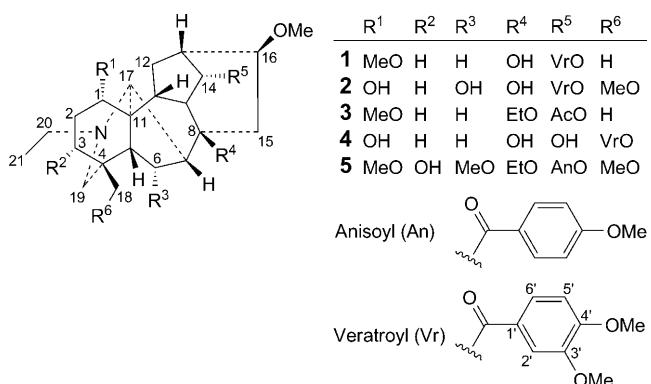


Fig. 1. The structures of compounds **1** – **5**

Results and Discussion. – Hemsleyaconitine A (**1**) was obtained as white prisms. Its molecular formula was determined to be $C_{32}H_{45}NO_7$ based on EI-MS (M^+ , m/z 555) and HR-ESI-MS (m/z 556.3287 ($[M + H]^+$; calc. 556.3274)) analyses, indicating eleven degrees of unsaturation. The IR spectrum showed the absorption for OH (3453 cm^{-1}), conjugated ester CO (1713 cm^{-1}), and aromatic ring (1602 , 1516 , 1463 cm^{-1}) functions.

The $^1\text{H-NMR}$ spectra of compound **1** displayed signals of one *N*-ethyl ($\delta(\text{H})$ 2.47–2.51 (*m*)), 2.48–2.54 (*m*), 1.07 (*t*, $J = 7.0$), two MeO groups ($\delta(\text{H})$ 3.20 (*s*), 3.29(*s*)), and one veratroyl group ($\delta(\text{H})$ 7.64 (*dd*, $J = 8.3$, 1.7), 7.57 (*d*, $J = 1.7$), 6.89 (*d*, $J = 8.4$), 3.92 (*s*), 3.93 (*s*)). Its $^{13}\text{C-NMR}$ (DEPT) spectrum revealed the presence of 32 C-atoms including six Me, seven CH_2 , and twelve CH groups, as well as seven quaternary C-atoms. These spectral data suggested that compound **1** might be an aconitine-type C_{19} -diterpenoid alkaloid [9][10]. Comparing the ^1H - and $^{13}\text{C-NMR}$ spectral data (*Tables 1* and 2) of compound **1** with those of geniculatine D [11] showed high similarity except that the signal due to C(18) was shifted upfield from $\delta(\text{C})$ 69.0 (CH_2) in geniculatine D to $\delta(\text{C})$ 26.4 (Me) in compound **1**, which suggested that there should be a Me(18) group located at C(4). The long-range correlations between Me(18) ($\delta(\text{H})$ 0.78 (*s*)) and C(3), C(4), C(5), and C(19) in the HMBC spectrum further confirmed the structure (*Fig. 2*).

Compound **1** had the same relative configuration as geniculatine D, not only being supported by their almost identical ^1H - and $^{13}\text{C-NMR}$ data (*Tables 1* and 2), but also verified by a ROESY spectrum (*Fig. 3*). As shown in *Fig. 2*, the correlations between H–C(10) (assumed to be in β -orientation with reference to the aconitine type C_{19} -diterpenoid alkaloids [2–4]), and H–C(1), H–C(9), H–C(13), and MeO–C(16) were observed, indicating the β -orientations of H–C(1), H–C(9), H–C(13), and MeO–C(16). Thus, the structure of compound **1** was determined as 18-dehydroxygeniculatine D, named hemsleyaconitine A (**1**).

Hemsleyaconitine B (**2**) was obtained as colorless prisms and had a molecular formula of $C_{32}H_{45}NO_9$ based on EI-MS (M^+ , m/z 587) and HR-ESI-MS (588.3173, $[M + H]^+$; calc. 588.3172) analyses. Careful interpretation of the NMR spectra suggested that compound **2** was also an aconitine type C_{19} -diterpenoid alkaloid. The ^1H - and $^{13}\text{C-NMR}$ data (*Tables 1* and 2) of compound **2** were almost identical to those of 14-*O*-veratroylneoline [12] except that the C-atom signal assignable to C(6) in compound **2** was shifted upfield at $\delta(\text{C})$ 72.0 from $\delta(\text{C})$ 83.2 in 14-*O*-veratroylneoline, implying that there was only one OH group located at C(6). This was further confirmed by the cross-peaks between H–C(6) ($\delta(\text{H})$ 4.67 (*d*, $J = 6.2$)), and C(4), C(5), C(7), and C(8) in the HMBC spectrum. Consequently, hemsleyaconitine B (**2**) was elucidated as 6-hydroxy-14-*O*-veratroylneoline.

Hemsleyaconitine C (**3**) was assigned the molecular formula $C_{27}H_{43}NO_5$ by analyses of EI-MS (M^+ , m/z 461) and HR-ESI-MS (462.3226 [$M + H]^+$; calc. 462.3219). The 1D-NMR spectra (*Tables 1* and 2) resembled those of 8-ethoxysachaconitine [13] with the exception of an additional AcO signal appearing at $\delta(\text{C})$ 171.4 (*s*), 21.4 (*q*). The AcO group was determined to be linked at C(14) by HMBCs between $\delta(\text{H})$ 4.71 (*t*, $J = 4.8$, H–C(14)) and $\delta(\text{C})$ 171.4 (C=O of the AcO group)). The ROESY correlations of H–C(14) with H–C(10) and H–C(13), together with the $^1\text{H}, ^1\text{H}$ -coupling constant similar to that of 8-ethoxysachaconitine, established the α -orientation of the AcO

Table 1. ^{13}C -NMR Data of Compounds **1–5**. At 125 MHz in CDCl_3 ; δ in ppm.

Position	1	2	3	4	5
C(1)	86.0 (<i>d</i>)	72.0 (<i>d</i>)	86.0 (<i>d</i>)	72.1 (<i>d</i>)	83.5 (<i>d</i>)
C(2)	26.7 (<i>t</i>)	29.2 (<i>t</i>)	27.0 (<i>t</i>)	26.7 (<i>t</i>)	33.0 (<i>t</i>)
C(3)	37.8 (<i>t</i>)	29.7 (<i>t</i>)	36.3 (<i>t</i>)	29.6 (<i>t</i>)	72.0 (<i>d</i>)
C(4)	34.5 (<i>s</i>)	37.8 (<i>s</i>)	34.3 (<i>s</i>)	36.9 (<i>s</i>)	42.9 (<i>s</i>)
C(5)	45.3 (<i>d</i>)	45.8 (<i>d</i>)	50.5 (<i>d</i>)	41.8 (<i>d</i>)	48.6 (<i>d</i>)
C(6)	25.4 (<i>t</i>)	72.9 (<i>d</i>)	24.5 (<i>t</i>)	25.1 (<i>t</i>)	82.7 (<i>d</i>)
C(7)	50.8 (<i>d</i>)	55.8 (<i>d</i>)	40.8 (<i>d</i>)	45.1 (<i>d</i>)	45.4 (<i>d</i>)
C(8)	74.0 (<i>s</i>)	75.6 (<i>s</i>)	77.6 (<i>s</i>)	74.1 (<i>s</i>)	78.3 (<i>s</i>)
C(9)	46.6 (<i>d</i>)	45.8 (<i>d</i>)	45.1 (<i>d</i>)	46.6 (<i>d</i>)	45.1 (<i>d</i>)
C(10)	45.3 (<i>d</i>)	43.7 (<i>d</i>)	43.2 (<i>d</i>)	44.0 (<i>d</i>)	45.1 (<i>d</i>)
C(11)	49.0 (<i>s</i>)	50.1 (<i>s</i>)	50.5 (<i>s</i>)	48.7 (<i>s</i>)	50.9 (<i>s</i>)
C(12)	28.6 (<i>t</i>)	29.3 (<i>t</i>)	29.1 (<i>t</i>)	28.4 (<i>t</i>)	28.7 (<i>t</i>)
C(13)	36.6 (<i>d</i>)	37.8 (<i>d</i>)	38.1 (<i>d</i>)	39.9 (<i>d</i>)	38.4 (<i>d</i>)
C(14)	76.6 (<i>d</i>)	76.6 (<i>d</i>)	75.8 (<i>d</i>)	75.7 (<i>d</i>)	75.9 (<i>d</i>)
C(15)	41.0 (<i>t</i>)	42.0 (<i>t</i>)	37.8 (<i>t</i>)	42.3 (<i>t</i>)	36.5 (<i>t</i>)
C(16)	81.8 (<i>d</i>)	82.1 (<i>d</i>)	83.4 (<i>d</i>)	81.8 (<i>d</i>)	83.2 (<i>d</i>)
C(17)	61.9 (<i>d</i>)	63.3 (<i>d</i>)	61.3 (<i>d</i>)	63.8 (<i>d</i>)	60.8 (<i>d</i>)
C(18)	26.4 (<i>q</i>)	80.4 (<i>t</i>)	26.5 (<i>q</i>)	70.0 (<i>t</i>)	76.8 (<i>t</i>)
C(19)	56.7 (<i>t</i>)	56.9 (<i>t</i>)	56.7 (<i>t</i>)	56.3 (<i>t</i>)	48.5 (<i>t</i>)
MeCH ₂ N	49.3 (<i>t</i>)	48.2 (<i>t</i>)	49.2 (<i>t</i>)	48.4 (<i>t</i>)	47.7 (<i>t</i>)
MeCH ₂ N	13.6 (<i>q</i>)	13.0 (<i>q</i>)	13.5 (<i>q</i>)	13.0 (<i>q</i>)	13.3 (<i>q</i>)
MeO–C(1)	56.3 (<i>q</i>)	—	56.2 (<i>q</i>)	—	55.7 (<i>q</i>)
MeO–C(6)	—	—	—	—	58.6 (<i>q</i>)
MeO–C(16)	56.0 (<i>q</i>)	56.2 (<i>q</i>)	56.2 (<i>q</i>)	56.3 (<i>q</i>)	56.3 (<i>q</i>)
MeO–C(18)	—	59.1 (<i>q</i>)	—	—	59.1 (<i>q</i>)
MeCH ₂ –C(8)	—	—	55.5 (<i>t</i>)	—	55.9 (<i>t</i>)
MeCH ₂ –C(8)	—	—	16.3 (<i>q</i>)	—	15.5 (<i>q</i>)
Acyl group	VrO–C(14) ^a	VrO–C(14)	AcO–C(14)	VrO–C(18)	AnO–C(14) ^b
C=O	166.3 (<i>s</i>)	165.9 (<i>s</i>)	171.4 (<i>s</i>)	166.3 (<i>s</i>)	166.2 (<i>s</i>)
C(1')	123.0 (<i>s</i>)	122.6 (<i>s</i>)	21.4 (<i>q</i>)	122.6 (<i>s</i>)	123.3 (<i>s</i>)
C(2')	112.0 (<i>d</i>)	112.1 (<i>d</i>)	—	112.0 (<i>d</i>)	131.7 (<i>d</i>)
C(3')	148.5 (<i>s</i>)	148.5 (<i>s</i>)	—	148.7 (<i>s</i>)	113.4 (<i>d</i>)
C(4')	152.8 (<i>s</i>)	152.9 (<i>s</i>)	—	153.1 (<i>s</i>)	163.1 (<i>s</i>)
C(5')	110.3 (<i>d</i>)	110.2 (<i>d</i>)	—	110.3 (<i>d</i>)	113.4 (<i>d</i>)
C(6')	123.5 (<i>d</i>)	123.5 (<i>d</i>)	—	123.4 (<i>d</i>)	131.7 (<i>d</i>)
MeO–C(3')	55.9 (<i>q</i>)	55.9 (<i>q</i>)	—	56.0 (<i>q</i>)	—
MeO–C(4')	56.0 (<i>q</i>)	56.0 (<i>q</i>)	—	56.0 (<i>q</i>)	55.3 (<i>q</i>)

^a) Vr = Veratroyl. ^b) An = Anisoyl.

group. Therefore, hemsleyaconitine C (**3**) was identified as 14-*O*-acetyl-8-ethoxysachaconitine.

The molecular formula of hemsleyaconitine D (**4**) was deduced as $\text{C}_{31}\text{H}_{43}\text{NO}_8$ from EI-MS (M^+ , m/z 557) and HR-ESI-MS (558.3076 [$M + \text{H}]^+$; calc. 558.3066) experiments. The ^1H - and ^{13}C -NMR spectra (*Tables 1* and *2*) of compound **4** were almost identical to those of columbianine [14] except for the presence of an additional veratroyl group ($\delta(\text{H})$ 7.62 (*dd*, $J = 8.4, 1.9$, H–C(6')), 7.51 (*d*, $J = 1.9$, H–C(2')), 6.88

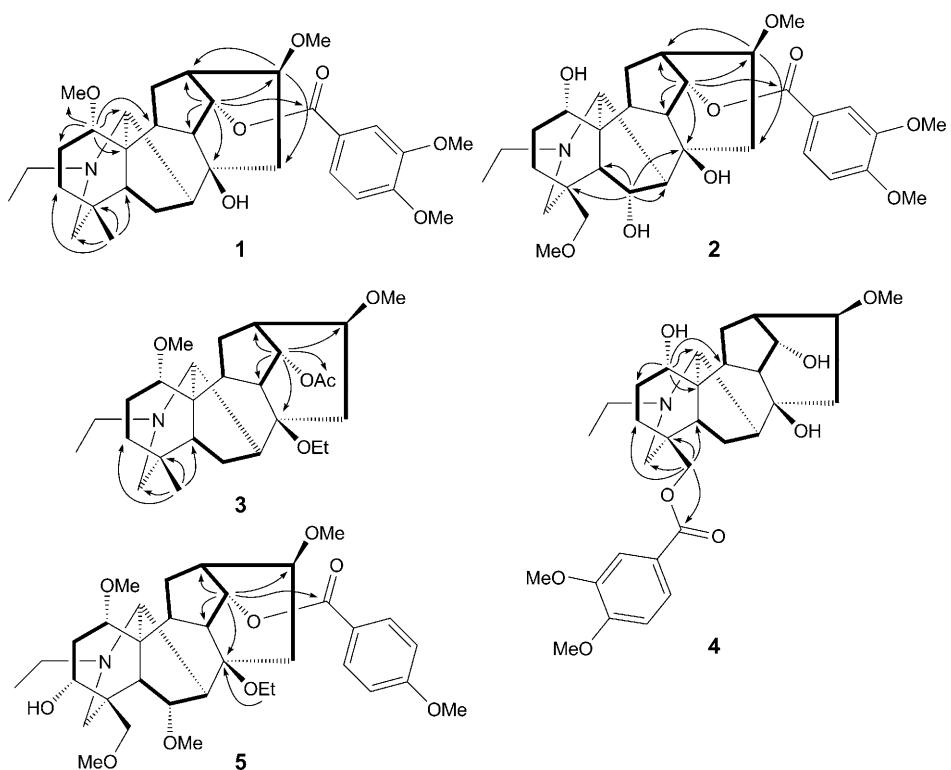
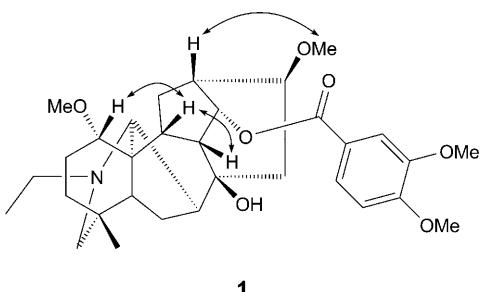
Table 2. $^1\text{H-NMR}$ Data of Compounds **1–5**. At 500 MHz in CDCl_3 ; δ in ppm, J in Hz.

Position	1	2	3	4	5
1	3.15 (<i>dd</i> , $J = 10.5, 6.6$) 1.96–2.00 (<i>m</i>)	3.70 (br. <i>s</i>) 1.83 (<i>dd</i> , $J = 14.0, 9.7$) 2.21–2.28 (<i>m</i>)	3.07 (<i>dd</i> , $J = 10.4, 6.7$) 1.93–1.98 (<i>m</i>) 2.22–2.29 (<i>m</i>)	3.75 (br. <i>s</i>) 1.76 (<i>dd</i> , $J = 14.0, 4.3$) 1.90–1.95 (<i>m</i>) 1.92–1.97 (<i>m</i>)	3.15 (<i>dd</i> , $J = 10.4, 6.2$) 1.81–1.86 (<i>m</i>) 2.30–2.35 (<i>m</i>) 3.81 (<i>dd</i> , $J = 12.3, 7.6$)
2a	2.32–2.37 (<i>m</i>)	—	—	—	—
2b	1.20–1.25 (<i>m</i>)	1.48–1.53 (<i>m</i>)	—	—	—
3a	1.23–1.27 (<i>m</i>)	1.59–1.66 (<i>m</i>)	2.11 (<i>dd</i> , $J = 14.2, 9.0$)	1.64–1.68 (<i>m</i>)	—
3b	1.45 (<i>d</i> , $J = 7.1$)	1.96 (<i>d</i> , $J = 5.7$)	1.34 (<i>d</i> , $J = 7.3$)	1.63–1.66 (overlapped)	2.39 (<i>d</i> , $J = 7.2$)
5	1.51 (<i>dd</i> , $J = 14.9, 8.1$) 1.88–1.94 (<i>m</i>)	4.67 (<i>d</i> , $J = 6.2$) —	1.28–1.36 (<i>m</i>) 1.92–1.98 (<i>m</i>)	1.61–1.66 (<i>m</i>) 1.92–1.98 (<i>m</i>)	4.13 (<i>d</i> , $J = 6.3$) —
6a	2.10 (br. <i>d</i> , $J = 7.8$)	1.90 (br. <i>s</i>)	2.34–2.37 (overlapped) 2.34–2.38 (overlapped)	1.88–1.93 (overlapped) 2.20–2.23 (overlapped)	2.47–2.49 (overlapped) 1.93–1.95 (overlapped)
6b	1.89–1.92 (overlapped)	2.35–2.39 (overlapped)	1.83–1.89 (<i>m</i>)	2.07–2.11 (<i>m</i>)	2.10–2.15 (<i>m</i>)
7	2.43–2.49 (<i>m</i>)	2.06–2.10 (<i>m</i>)	1.92–1.99 (<i>m</i>)	1.59–1.65 (<i>m</i>)	1.92–1.95 (<i>m</i>)
9	1.95–1.99 (<i>m</i>)	1.57–1.64 (<i>m</i>)	1.83–1.89 (<i>m</i>)	2.00–2.05 (<i>m</i>)	2.14–2.19 (<i>m</i>)
10	2.25 (<i>dd</i> , $J = 15.0, 5.9$)	2.11–2.17 (<i>m</i>)	2.34–2.39 (overlapped)	2.31 (<i>dd</i> , $J = 7.5, 5.4$)	2.45–2.51 (overlapped)
12a	2.63 (<i>dd</i> , $J = 6.7, 5.4$)	2.59 (<i>dd</i> , $J = 6.2, 5.8$)	5.09 (<i>t</i> , $J = 4.3$)	4.71 (<i>t</i> , $J = 4.8$)	4.96 (<i>t</i> , $J = 4.8$)
12b	5.14 (<i>t</i> , $J = 4.7$)	2.02–2.07 (<i>m</i>)	2.06–2.12 (<i>m</i>)	1.17–1.23 (<i>m</i>)	2.10–2.15 (<i>m</i>)
13	2.02–2.04 (overlapped)	2.27–2.31 (<i>m</i>)	1.57 (<i>d</i> , $J = 11.3$)	2.38 (<i>dd</i> , $J = 12.9, 9.2$)	2.13–2.19 (<i>m</i>)
14	3.27–3.30 (overlapped)	3.27–3.37 (overlapped)	3.21–3.24 (overlapped)	3.36 (<i>dd</i> , $J = 9.1, 5.1$)	3.31–3.34 (overlapped)
15a	2.99 (br. <i>s</i>)	2.68 (br. <i>s</i>)	2.73 (br. <i>s</i>)	2.81 (br. <i>s</i>)	2.66 (br. <i>s</i>)
15b	0.78 (<i>s</i>)	3.30–3.35 (<i>m</i>)	0.73 (<i>s</i>)	3.95 (<i>d</i> , $J = 10.8$)	3.41 (<i>d</i> , $J = 8.7$)
16	3.64 (<i>d</i> , $J = 8.4$)	2.31–2.35 (overlapped)	—	4.11 (<i>d</i> , $J = 10.8$)	3.57 (<i>d</i> , $J = 8.6$)
17	2.00–2.04 (overlapped)	2.74 (<i>d</i> , $J = 10.6$)	1.93–1.97 (<i>m</i>)	2.05–2.11 (<i>m</i>)	3.31–3.35 (overlapped)
18a	2.41–2.46 (<i>m</i>)	2.51 (<i>dd</i> , $J = 12.8, 7.1$), 2.48–2.54 (<i>m</i>)	2.38–3.46 (<i>m</i>)	2.44 (<i>d</i> , $J = 11.0$) 2.30–2.36 (<i>m</i>), 2.41–2.45 (<i>m</i>)	3.45 (<i>d</i> , $J = 11.1$) 2.39–2.45 (<i>m</i>), 2.57 (<i>dd</i> , $J = 12.4, 7.2$), 1.13 (<i>t</i> , $J = 7.2$)
18b	—	1.14 (<i>t</i> , $J = 7.0$)	1.03 (<i>t</i> , $J = 7.1$)	1.07 (<i>t</i> , $J = 7.1$)	1.07 (<i>t</i> , $J = 7.1$)
19a	2.47–2.52 (<i>m</i>)	—	3.24 (<i>s</i>)	—	3.24 (<i>s</i>)
19b	2.47–2.51 (<i>m</i>), 2.48–2.54 (<i>m</i>)	2.53 (<i>dd</i> , $J = 12.9, 7.1$)	3.27 (<i>s</i>)	3.33 (<i>s</i>)	3.33 (<i>s</i>)
MeCH_2N	1.07 (<i>t</i> , $J = 7.0$)	—	—	—	3.29 (<i>s</i>)
MeO-C(1)	3.20 (<i>s</i>)	—	3.29–2.36 (<i>m</i>)	—	2.85–2.88 (<i>m</i>)
MeO-C(6)	—	—	1.07 (<i>t</i> , $J = 6.9$)	—	0.80 (<i>t</i> , $J = 6.9$)
MeO-C(16)	3.29 (<i>s</i>)	—	—	—	—
MeO-C(18)	—	3.31 (<i>s</i>)	—	—	—
$\text{MeCH}_2\text{-C(8)}$	—	—	—	—	—
$\text{MeCH}_2\text{-C(8)}$	—	—	—	—	—

Table 2 (cont.)

Position	1	2	3	4	5
Acyl group	VrO-C(14) ^a)	VrO-C(14)	AcO-C(14)	VrO-C(18)	AnO-C(14) ^b)
1'	–	–	2.00 (s)	–	–
2'	7.57 (<i>d, J</i> = 1.7)	7.59 (s)	–	7.51 (<i>d, J</i> = 1.9)	8.02 (<i>d, J</i> = 8.7)
3'	–	–	–	–	6.89 (<i>d, J</i> = 8.8)
5'	6.89 (<i>d, J</i> = 8.4)	6.87 (<i>d, J</i> = 8.4)	–	6.88 (<i>d, J</i> = 8.5)	8.02 (<i>d, J</i> = 8.7)
6'	7.64 (<i>dd, J</i> = 8.3, 1.7)	7.65 (<i>d, J</i> = 8.3)	–	7.62 (<i>dd, J</i> = 8.4, 1.9)	6.89 (<i>d, J</i> = 8.8)
MeO-C(3')	3.92 (s)	3.91 (s)	–	3.91 (s)	–
MeO-C(4')	3.93 (s)	3.92 (s)	–	3.92 (s)	3.84 (s)

^a) Vr = Veratroyl. ^b) An = Anisoyl.

Fig. 2. Selected ^1H , ^1H -COSY (—) and HMBC ($\text{H}\rightarrow\text{C}$) correlations of compounds **1–5**Fig. 3. Selected ROESY correlations of compound **1**

($d, J=8.5$, $\text{H}-\text{C}(5')$), 3.91 (s , $\text{MeO}-\text{C}(3')$), 3.92 (s , $\text{MeO}-\text{C}(4')$) in compound **4**. The veratroyl group at $\text{C}(18)$ was evidenced by the HMBC correlations from $\text{H}_a-\text{C}(18)$ and $\text{H}_b-\text{C}(18)$ to $\text{C}=\text{O}$ of the veratroyl group. Accordingly, hemsleyaconitine D (**4**) was established as 18-veratroylkaracoline (Fig. 1).

Hemsleyaconitine E (**5**) had the molecular formula $\text{C}_{35}\text{H}_{51}\text{NO}_9$, in agreement with the EI-MS (M^+ , m/z 629) and HR-ESI-MS (positive-ion mode; 630.3649 ($[M+\text{H}]^+$;

calc. 630.3642)) analyses. The ^1H - and ^{13}C -NMR spectra (*Tables 1* and *2*) differed from those of austroconitine B [15] in the substitution pattern of C(8), where an EtO group ($\delta(\text{H})$ 2.85–2.88 (*m*, 2 H), 0.80 (*t*, J = 6.9, 3 H)) was discernible, instead of a OH group in austroconitine B. This contention was also confirmed by the HMBC between $\delta(\text{H})$ 2.85–2.88 (EtO) and C(8). Hence, hemsleyaconitine E (**5**) was determined as 8-*O*-ethyl austroconitine B.

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Experimental Part

General. Column chromatography (CC): silica gel (SiO₂; 200–300 mesh, *Qingdao Marine Chemical Ltd.*, Qingdao, P. R. China); Al₂O₃ (*Shanghai Wusi Chemical Reagents Company, Ltd.*); Sephadex LH-20 (*Pharmacia Fine Chemical Co. Ltd.*, Germany). M.p.: *XRC-1* micro-melting-point apparatus, uncorrected. Optical rotations: *Horiba SEPA-300* polarimeter. UV Spectra: *Shimadzu UV-2401A* spectrophotometer. IR Spectra: *Bio-Rad FTS-135* spectrometer. 1D- and 2D-NMR spectra: *Bruker AM-400* and *DRX-500* spectrometers; chemical shifts δ in ppm with reference to the solvent signals. MS: *VG Autospec-3000* spectrometer at 70 eV; in *m/z*. HR-ESI-MS: *API Qstar-Pulsar-1* spectrometer.

Plant Material. The roots of *Aconitum hemsleyanum* PRITZ. were collected in Wuding County, Yunnan Province, P. R. China, in October, 2006, and authenticated by Prof. Dr. Li-Gong Lei from Kunming Institute of Botany. A voucher specimen (No. KIB 2006-10-01) has been deposited with the State Key Laboratory of Phytochemistry and Plant Resources in West China, Kunming Institute of Botany, Chinese Academy of Sciences.

Extraction and Isolation. The roots of *A. hemsleyanum* (54 kg) were powdered and extracted three times with 95% EtOH for 2 h under reflux. After removing the solvent, the crude extract was dissolved in 15 l of 2% HCl soln. and filtered. The acidic soln. was basified to pH 9.0 with NH₃ (25%) and then extracted with CHCl₃ to obtain crude alkaloid extract (460 g) after removal of CHCl₃ *in vacuo*. The extract was chromatographed over a SiO₂ column (4.6 kg, 200–300 mesh) and eluted with gradient petroleum ether (PE)/acetone/Et₂NH (10:1:1 → 5:1:1) to provide six fractions, *Frs. 1–6*. *Fr. 2* (14.6 g) was successively chromatographed on a SiO₂ column (PE/acetone/Et₂NH, 15:1:1), an Al₂O₃ column (PE/acetone, 6:1), and *Sephadex LH-20* (CHCl₃/MeOH, 1:1) to yield **1** (12 mg), **3** (20 mg), and sachaconitine [5] (67 mg). *Fr. 3* (30.3 g) was subjected to a SiO₂ column and eluted with PE/acetone/Et₂NH 15:3:1, followed by Al₂O₃ CC (PE/acetone 5:1) to yield **4** (18 mg), karakoline [6] (2.5 g), and isotalatizidine [8] (1.8 g). *Fr. 4* (42.0 g) was subjected to an Al₂O₃ column with the eluent of PE/acetone 4:1 to yield **2** (15 mg), **5** (25 mg), and acorestine [7] (800 mg).

Hemsleyaconitine A (=18-Dehydroxygeniculatine D = (1*a*,14*a*,16*β*)-20-Ethyl-8-hydroxy-1,16-dimethoxy-4-methylaconitan-14-yl 3,4-Dimethoxybenzoate; **1**). White prisms. M.p. 91–92°. $[\alpha]_{\text{D}}^{25.1} = +21.38$ (*c* = 0.11, CHCl₃). UV (MeOH): 219 (2.26). IR (KBr): 3453, 2963, 2928, 1713, 1602, 1516, 1463, 1270, 1222, 1094, 1025, 765. NMR: *Tables 1* and *2*. EI-MS: 555 (1, M^+), 524 (100, $[M - \text{MeO}]^+$), 262 (33), 165 (100). HR-ESI-MS (pos.): 556.3287 ($[M + \text{H}]^+$; C₃₂H₄₆NO₅⁺; calc. 556.3274).

Hemsleyaconitine B (=6-Hydroxy-14-O-veratroylneoline = (1*a*,6*a*,14*a*,16*β*)-20-Ethyl-1,6,8-trihydroxy-16-methoxy-4-(methoxymethyl)aconitan-14-yl 3,4-Dimethoxybenzoate; **2**). Colorless prisms. M.p. 104–105°. $[\alpha]_{\text{D}}^{24.9} = +9.16$ (*c* = 0.13, CHCl₃). UV (MeOH): 219 (2.37). IR (KBr): 3423, 2965, 2933, 1713, 1602, 1516, 1464, 1271, 1223, 1109, 1041, 764. NMR: *Tables 1* and *2*. EI-MS: 587 (4, M^+), 570 (39, $[M - \text{OH}]^+$), 285 (10), 165 (100). HR-ESI-MS (pos.): 588.3173 ($[M + \text{H}]^+$, C₃₂H₄₆NO₅⁺; calc. 588.3172).

Hemsleyaconitine C (=14-O-Acetyl-8-ethoxysachaconitine = (1*a*,14*a*,16*β*)-8-Ethoxy-20-ethyl-1,16-dimethoxy-4-methylaconitan-14-yl Acetate; **3**). Colorless crystals. M.p. 133–134°. $[\alpha]_{\text{D}}^{25.8} = -31.98$ (*c* = 0.17, CHCl₃). UV (MeOH): 223 (1.45). IR (KBr): 3449, 2969, 2924, 2891, 1739, 1640, 1461, 1443, 1365, 1248, 1117, 1094. NMR: *Tables 1* and *2*. EI-MS: 461 (4, M^+), 430 (47, $[M - \text{MeO}]^+$), 402 (80), 131 (100), 91 (85), 71 (98). HR-ESI-MS (pos.): 462.3226 ($[M + \text{H}]^+$, C₂₇H₄₄NO₅⁺; calc. 462.3219).

Hemsleyaconitine D ($=18$ -Veratroylkaracoline [$(1\alpha,14\alpha,16\beta)$ - 20 -Ethyl- $1,8,14$ -trihydroxy- 16 -methoxyaconitan- 4 -yl]methyl $3,4$ -Dimethoxybenzoate; **4**). White prisms. M.p. 102 – 103° . $[\alpha]_D^{24,3} = +13.16$ ($c = 0.30$, CHCl_3). UV (MeOH): 219 (2.33). IR (KBr): 3426, 2936, 2837, 1711, 1601, 1516, 1463, 1418, 1291, 1271, 1222, 1176, 1101, 1023, 763. NMR: *Tables 1* and *2*. EI-MS: 557 (12, M^+), 540 (63, $[M - \text{OH}]^+$, 319 (29), 165 (100). HR-ESI-MS (pos.): 558.3076 ($[M + \text{H}]^+$, $\text{C}_{31}\text{H}_{44}\text{NO}_8^+$; calc. 558.3066).

Hemsleyaconitine E ($=8$ -O-Ethylaustroconitine *B* [$(1\alpha,3\alpha,6\alpha,14\alpha,16\beta)$ - 8 -Ethoxy- 20 -ethyl- 3 -hydroxy- $1,6,16$ -trimethoxy- 4 -(methoxymethyl)aconitan- 14 -yl 4-Methoxybenzoate; **5**). White prisms. M.p. 93 – 94° . $[\alpha]_D^{24,2} = +13.89$ ($c = 0.19$, CHCl_3). UV (MeOH): 257 (2.13). IR (KBr): 3457, 2970, 2930, 2890, 1713, 1607, 1512, 1460, 1293, 1278, 1169, 1119, 1098, 771. NMR: *Tables 1* and *2*. EI-MS: 629 (2, M^+), 598 (100, $[M - \text{MeO}]^+$), 135 (59). HR-ESI-MS (pos.): 630.3649 ($[M + \text{H}]^+$, $\text{C}_{35}\text{H}_{52}\text{NO}_9^+$; calc. 630.3642).

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